# **Bioefficiency of Botanicals against** *Colletotrichum truncatum*, Causing Pod Blight of Soybean

Chavan S.S.<sup>1</sup> and Suryawanshi A.P.<sup>2</sup>

<sup>1</sup>Department of Plant Pathology, Sau. K. S K College of Agriculture, Beed, VNMKV, Parbhani <sup>2</sup>Department of Plant Pathology, College of Agriculture, VNMKV, Parbhani E-mail: <sup>1</sup>smeetachavan4@gmail.com, <sup>2</sup>smeetachavan4@gmail.com

Abstract—Soybean (Glycine max L. Merrill.) is one of the most important oilseed-cum leguminous crop, which is the largest source of vegetable oil and protein in the world. Colletotrichum truncatum has been reported as major constraint in the successfully cultivation of soybean, causing more than 30 per cent yield losses. A total of eleven botanicals viz., Mehandi (Lawsonia innermis), Ginger (Zingiber officinale), Parthenium (Parthenium hysterophorus), Neem (Azardirachta indica), Garlic (Allium sativum), Turmeric (Curcuma longa), Bougainvillea (Bougainvillea spectabilis), Onion (Allium cepa), Eucalyptus (Eucalyptus globules), Datura (Datura metal), Beshram (Ipomea carnea) were evaluated (@ 10, 15 and 20% each) in vitro against C. truncatum, applying poisoned food technique (Nene and Thapliyal, 1993).

All the botanicals/plant leaf extracts tested in vitro were found significantly effective in reducing the percentage mycelial growth of C. truncatum over untreated control. The mean radial mycelial growth recorded with the plant extracts tested (@ 10, 15 and 20% each) was ranged from 14.68 mm (Garlic) to 54.94 mm (Beshram). Garlic recorded lowest mean colony diameter (14.68 mm) and highest mean mycelial growth inhibition (83.69%) of the test pathogen over untreated control. This was followed by the botanicals, onion (Mean Col. Dia. 32.32 mm and mean inhibition 64.09%), Ginger (Mean Col. Dia. 38.66 mm and mean inhibition 57.05%), Neem (Mean Col. Dia. 43.52 mm and mean inhibition 51.64%), Parthenium (Mean Col. Dia. 45.78 mm and mean inhibition 49.13%), turmeric (Mean Col. Dia. 50.94 mm and mean inhibition 43.39%). Bougainvillea (Mean Col. Dia. 51.98 mm and mean inhibition 42.69%), Eucalyptus (Mean Col. Dia. 52.60 mm and mean inhibition 41.56%) and Mehandi (Mean Col. Dia. 53.29 mm and mean inhibition 40.79%). The least effective found were Datura (Mean Col. Dia. 54.37 mm and mean inhibition 39.59%) and Beshram (Mean Col. Dia. 54.94 mm and mean inhibition 38.95%).

**Keywords**: Colletotrichum truncatum, mean inhibition, fungistatic and botanicals.

# 1. INTRODUCTION

Soybean is the world's foremost provider of protein and oil. In Maharashtra, the area production and productivity of soybean were 32.13 lakh hectare, 39.95 lakh metric tonnes and 1243 kg/ha, respectively [1]. Soybean growing major states in the country are Madhya Pradesh, Maharashtra, Karnataka, Andhra Pradesh. Soybean plant health is a critical component of

profitable soybean production. *Colletotrichum truncatum*, is the most common species recorded on soybean [9] and the crop soybean is susceptible to *C. truncatum* at all stages of development particularly from bloom to pod fill.

### 2. MATERIAL AND METHODS

### **Preparation of plant extracts**

Hundred grams of fresh healthy plant parts (leaves/root/bulbs) collected from field were washed with distilled water and airdried and crushed in 100 ml of distilled water (w/v). The crushed product was filtered through double layer, muslin cloth and further filtrated through Whatsman No. 1 filter paper using funnel and volumetric flasks (100 ml cap.). The prepared solution gave 100 per cent concentration, which was further diluted to required concentrations of 10.0, 15.0 and 20.0 per cent. The extracts were tested against *C. truncatum* on the cultural media using poison food technique [11] under *in vitro* condition.

An appropriate quantity of each plant extract (100%) was separately mixed thoroughly with autoclaved and cooled  $(40^{\circ}\text{C})$  PDA medium in conical flasks (250 ml cap.) to obtain desired concentrations (10, 15 and 20 per cent). The PDA medium amended separately with plant extract was then poured (20 ml/plate) into sterile glass Petri plates (90 mm dia.) and allowed to solidify at room temperature. For each test botanical extract and their respective concentrations, three plates / treatment / replication were maintained. Each plant extract and its respective concentrations were replicated thrice. Upon solidification of PDA, all the treatment and control plates were aseptically inoculated by placing in the centre a 5 mm mycelial disc obtained from a week old actively growing pure culture of C. truncatum. Plates containing plain PDA without any botanical extract and inoculated with mycelial disc of the test fungus served as untreated control. All these plates were then incubated at  $27 \pm 2^{\circ}C$  temperature for a week or till the untreated control plates were fully covered with mycelial growth of the test fungus.

Details of the ex		
Design :	CRD	
Replications	:	Two
Treatments	:	12

		Local name	Scientific Name
T <sub>1</sub>	:	Mehandi	Lawsonia innermis
T <sub>2</sub>	:	Ginger	Zingiber officinale
T <sub>3</sub>	:	Parthenium	Parthenium hysterophorus
$T_4$	:	Neem	Azardirachta indica
T <sub>5</sub>	:	Garlic	Allium sativum
T <sub>6</sub>	:	Turmeric	Curcuma longa
T <sub>7</sub>	:	Bougainvillea	Bougainvillea spectabilis
T <sub>8</sub>	:	Onion	Allium Cepa
T <sub>9</sub>	:	Eucalyptus	Eucalyptus globulus
T <sub>10</sub>	:	Datura	Datura metal
T <sub>11</sub>	:	Beshram	Ipomea carnea
T <sub>12</sub>	:	Control	
		(untreated)	

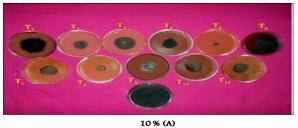
Observations on radial mycelial growth of the test fungus were recorded at 24 hrs. interval and continued till growth of the test pathogen in untreated control plate was fully covered. Per cent inhibition of the test pathogen was calculated by applying the formula [16]. Observations on sporulation were recorded at 10 days after incubation.

## 3. RESULTS AND DISCUSSION

The results revealed that (Table No. 1 and plate I) all the botanicals/plant leaf extracts tested in vitro were found significantly effective in reducing the percentage mycelial growth of C. truncatum over untreated control. The mean radial mycelial growth recorded with the plant extracts tested (@ 10, 15 and 20% each) was ranged from 14.68 mm (Garlic) to 54.94 mm (Beshram). Garlic recorded lowest mean colony diameter (14.68 mm) and highest mean mycelial growth inhibition (83.69%) of the test pathogen over untreated control. This was followed by the botanicals, onion (Mean Col. Dia. 32.32 mm and mean inhibition 64.09%), Ginger (Mean Col. Dia. 38.66 mm and mean inhibition 57.05%), Neem (Mean Col. Dia. 43.52 mm and mean inhibition 51.64%), Parthenium (Mean Col. Dia. 45.78 mm and mean inhibition 49.13%), turmeric (Mean Col. Dia. 50.94 mm and mean inhibition 43.39%), Bougainvillea (Mean Col. Dia. 51.98 mm and mean inhibition 42.69%), Eucalyptus (Mean Col. Dia. 52.60 mm and mean inhibition 41.56%) and Mehandi (Mean Col. Dia. 53.29 mm and mean inhibition 40.79%). The least effective found were Datura (Mean Col. Dia. 54.37 mm and mean inhibition 39.59%) and Beshram (Mean Col. Dia. 54.94 mm and mean inhibition 38.95%). Botanicals garlic, onion, ginger, neem, mehandi, parthnium, bougainvillea were also reported fungistatic against several Colletotrichum species causing anthracnose, blights, leaf spot in many crop by several workers [6,10, 4, 2, 15, 3, 12, 13, 5, 8, 14 and 7].

Table 1 : Bioefficacy of botanicals against C. truncatum										
Treatments	Col.	dia.*(	mm)	Av.	% Inhibition			Av.		
	at Conc.			(m	/			(%)		
	10	15	20	m)	10 %	15 %	20 %	~ /		
	%	%	%	,						
Lawsonia	56.2	53.5	50.1	53.2	37.48	40.55	44.33	40.79		
innermis	7	0	0	9	(22.0	(23.9	(26.3	(24.0		
(Mehandi)					0)	1)	1)	7)		
Zingiber	42.1	38.6	35.2	38.6	53.15	57.11	60.89	57.05		
officinale	7	0	0	6	(32.0	(34.8	(37.5	(34.8		
(Ginger)					9)	0)	1)	0)		
Parthenium	51.3	44.2	41.8	45.7	42.96	50.89	53.55	49.13		
hysterophor	3	0	0	8	(25.4	(30.5	(32.3	(29.4		
us					3)	6)	7)	5)		
(Parthenium										
)										
Azardiracht	46.6	43.6	40.3	43.5	48.16	51.55	55.22	51.64		
a indica	7	0	0	2	(28.7	(31.0	(33.5	(31.1		
(Neem)					6)	1)	8)	2)		
Allium	18.3	14.5	11.2	14.6	79.63	83.88	87.55	83.69		
sativum	3	0	0	8	(52.7	(57.0	(61.1	(56.9		
(Garlic)					7)	1)	0)	6)		
Curcuma	55.2	50.3	47.3	50.9	38.63	44.11	47.44	43.39		
longa	3	0	0	4	(22.7	(20.1	(28.3	(23.7		
(Turmeric)					1)	5)	2)	3)		
Bougainville	55.2	51.7	47.7	51.5	38.59	42.47	47.00	42.69		
a spectabilis	7	7	0	8	(22.6	(25.1	(28.0	(25.2		
(Bougainvill					9)	2)	2)	8)		
ea)										
Allium cepa	35.1	32.1	29.7	32.3	60.93	64.33	67.00	64.09		
(Onion)	7	0	0	2	(37.5	(40.0	(42.0	(39.8		
					3)	0)	6)	6)		
Eucalyptus	56.0	52.1	49.7	52.6	37.78	42.11	44.78	41.56		
globules	0	0	0	0	(22.1	(24.8	(26.6	(24.5		
(Eucalyptus)					8)	8)	0)	5)		
Datura	57.1	54.6	51.4	54.3	36.55	39.33	42.89	39.59		
metal	0	0	0	7	(21.4	(23.1	(25.3	(23.3		
(Datura)					3)	3)	8)	1)		
Ipomea	57.3	54.8	52.7	54.9	36.30	39.11	41.44	38.95		
carnea	3	0	0	4	(21.2	(23.0	(24.4	(22.9		
(Beshram)					8)	1)	9)	3)		
Untreated	90.0	90.0	90.0	90.0						
(Control)	0	0	0	0						
S.E. <u>+</u>	0.05	0.03	0.06		0.63	0.63	0.66			
C.D.	0.15	0.10	0.18		1.87	1.84	1.93			
(P=0.05)										
	* :- Average of four replication, Av. :- Average, % :- per cent,									
dia.:- Diameter										

### PLATE I





15% (B)

20 % (C)

# In vitro effect of botanicals at (A) 10 %, (B) 15 % and (C) 20 % on growth and inhibition of *C. truncatum*

- T<sub>1</sub>: Lawsonia innermis T<sub>2</sub>: Zingiber officinale T<sub>3</sub>: Parthenium hysterophorus
- r<sub>3</sub> : Farmentum nysterop r<sub>4</sub> : Azardirachta indica r<sub>5</sub> : Allium sativum r<sub>6</sub> : Curcuma longa

 $\begin{array}{l} T_7: Bougainvillea spectabilis\\ T_8: Allium cepa\\ T_9: Eucalyptus globulus\\ T_{10}: Datura metal\\ T_{11}: Ipomea carnea\\ T_{12}: Untreated control \end{array}$ 

### REFERENCES

- 1]Anonymous, (2012). The Soybean Processors Association of India, SOPA, Indore (MP) India.
- 2] Chandrasekaran, A. and Rajappan, K. (2002). Effect of plant extracts, antagonists and chemicals (individual and combined) on foliar anthracnose and pod blight of soybean. *J. Mycol. Pl. Patho.*, **32** (1): 25-27.
- 3] George, A.P., Rani, S.V. and Sivam, S.M. (2003). *In vitro* effect of some plant extracts against Chilli anthracnose. *Indian Phytopath.*, **56** (3): 317.
- 4] Gomathi, V. and Kannabiran, B. (2000). Inhibitory effects of leaf extracts of some plants on the anthracnose fungi infecting *Capsicum annum. Indian Phytopath.* **53** (3) : 305-308.

- 5] Gorawar, M.M., Hedge, V.R. and Kulkarni, S. (2006). Biology and management of leaf spot of turmeric caused by *Colletotrichum capsici. J. Pl. Dis. Sci.*, **1** (2): 156-158.
- 6] Gupta, J.S., Agarwal, M.B., Dixit, R.B. and Agarwal, M. (1981). Effect of metabolites from different host plants on conidial germination of *Colletotrichum graminicola* and *Colletotrichum capsici. Geobios*, **8** : 226-228.
- 7] Jat, B.L., Gour, H.N. and Sharma, P. (2008). Efficacy of phytoextracts and bioagents against *Collectotrichum gloeosporiodes* causing banana fruit rot. J. Mycol. Pl. Pathol., 38(3): 635-638.
- 8] Kumar, S. and Yadav, B.P. (2007). Efficacy of fungicides and phytoextract on *Colletotrichum* spp. J. Mycol. Pl. Pathol., 37(2): 363-364.
- 9] Lenne, J.M. (1992). *Colletotrichum* disease of legumes. In: *Colletotrichum:* biology, pathology and control. (Eds.J.Bailey and M.J. Jeger), Red Wood Press Ltd. Melksham, U.K. PP: 134-136.
- 10] Mesta, R.K. (1996). Studies on fruit rot of chilli (*Capsicum annum* L.) caused by *Colletotrichum capsici* (Sydow) Butler and Bisby. *M.Sc. (Agri.) Thesis*, Univ. Agric. Sci., Dharwad, Karnataka, India.
- 11] Nene, Y.L.; Thapliyal, P.N.; Srivastava, S.S.L; Sarbhoy, A.K. and Khare, M.N. (1972). Seed and seedling rots of soybean. *Fung. and Nem. Tests.* **28** : 266.
- 12] Rangarajulu, K.; Gomathi, V. and Kannabiran, B. (2003). Fungitoxic effect of root extracts of certain plant species on *C.capsici*, causing anthracnose in *Capsicum annuum*. *Indian Phytopath.* **56** (1): 114-116.
- 13] Rao, C.H. and Narayana, Y.D. (2005). In vitro evaluation of fungicides, plant extracts and biocontrol agents against C. dematum (Pers. Ex. Fr.) Grove the causal organism of chickpea (Cicer arietenum L.) blight. In national symposium on crop disease management in dry land Agril. and 57<sup>th</sup> Annual meeting IPS, Jan. 12-14, 2005, MAU, Parbhani.
- 14] Sharma, A., Dass, A. and Paul, M.S. (2007). Antifungal effect of neem extract on some common phytopathogenic fungi. *Adv. in Pl. Sci.* 20 (2): 357-358.
- 15] Swamy, B.S. and Kulkarni, S. (2003). *In-vitro* evaluation of fungicides and botanicals against *C. capsici*, causing leaf spot of Turmeric. *Indian Phytopath*, 56(3): 339-340.
- 16] Vincent, J.M. (1927). Distortion of fungal hyphae in the presence of certain inhibitors Nature : 159-180.